

PARASITOIDS ASSOCIATED WITH THE MACROLEPIDOPTERA
COMMUNITY AT COOPER'S ROCK STATE FOREST, WEST VIRGINIA:
A BASELINE STUDY¹

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Abstract.—During 1984 and 1985 macrolepidopterous larvae were collected by means of pole pruning from foliage of black birch, mixed oaks, black cherry, and red maple at Cooper's Rock State Forest in northern West Virginia. From 46 species of caterpillars, 74 species of parasitoids in eight families were reared. The most abundant parasitoids were Tachinidae (Diptera): *Hyphantrophaga virilis* (Aldrich and Webber) and *Compsilura concinnata* (Meigen); Braconidae (Hymenoptera): *Protapanteles paleacritae* (Riley), *Microplitis hyphantriae* (Ash.), and *Diolcogaster facetosa* (Weed); Eulophidae (Hymenoptera): *Euplectrus maculiventris* (Westwood), *Eulophus anomocerus* (Crawford), and *E. nebulosus* (Prov.); and Perilampidae (Hymenoptera): *Perilampus canadensis* (Crawford).

Key Words: Parasitoids, macrolepidoptera community, baseline study

In 1984 and 1985, Cooper's Rock State Forest in northern West Virginia was just west of the leading edge of the infestation of the gypsy moth, *Lymantria dispar* (L.) (Lepidoptera: Lymantriidae). During those two years, a study was conducted to obtain baseline data for native lepidopterous defoliators and their parasitoids before the buildup of gypsy moth and subsequent defoliation and suppression efforts.

Previous reports have summarized hymenopterous and dipterous parasitoids associated with macrolepidopterous larvae (Muesebeck et al. 1951, Krombein and Burks 1967, Krombein 1958, Krombein et al. 1979, Viereck 1916, Arnaud 1978, Raizenne 1952, Schaffner and Griswold 1934, Townes and Townes 1959, 1960, 1962). Butler (1990) recorded 28 species of para-

sitoids associated with a "looper" complex in eastern West Virginia.

During the baseline study at Cooper's Rock State Forest, 400 species of adult macrolepidoptera were collected by blacklight trap (Butler and Kondo 1991) and 100 species of macrolepidopterous larvae were collected from the most abundant host trees (Butler 1992). Here I report results of the associated study of parasitoids reared from macrolepidoptera larvae.

METHODS AND MATERIALS

The West Virginia University Forest at Cooper's Rock State Forest is located at 561 m in Preston and Monongalia counties about 32 km east of Morgantown, West Virginia. The area consists of a 50- to 60-year-old mixed mesophytic forest (Carvell 1983). The most abundant tree species in the study area are red maple (*Acer rubrum* L., Aceraceae), white and red oak (*Quercus alba* L., *O. rubra* L., Fagaceae), black cherry (*Prunus serotina*

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Ehrh., Rosaceae), and black birch (*Betula lenta* L., Betulaceae).

Macrolepidopterous larvae were sampled from the mid- and lower canopies of red maple, mixed oaks, black cherry and black birch using pole pruners with large plastic catch bags. Samples were taken once each week from 16 May to 11 October 1984 and from 2 May to 3 October 1985.

In the laboratory, macrolepidopterous larvae were removed from the foliage, identified and placed for rearing in 150 × 25 ml plastic Petri dishes on bouquets of fresh foliage of the appropriate host plant. At pupation, Lepidoptera were placed in jars of moist vermiculite and held at 24° C or, for those species requiring a cold period, jars were held at 4° C for a minimum of 90 days. As parasitoid larvae emerged and pupated, they were placed in 4 dram vials of moist vermiculite plugged with cotton. Adults emerging from pupation vials or directly from macrolepidopterous larvae or pupae were killed, pinned, labeled and sent to specialists for identification. Voucher specimens are located in the West Virginia University Arthropod Collection.

RESULTS

Parasitoids were reared from 46 Macrolepidoptera species (Table 1). A total of 74 parasitoid species in eight families was recorded. The number of parasitoid species in each family is as follows: 11 Tachinidae, 29 Ichneumonidae, 16 Braconidae, 1 Trigonidae, 1 Chalcididae, 13 Eulophidae, 1 Pteromalidae and 2 Perilampidae.

Species reared most frequently from four or more different host species include Tachinidae *Hyphantrophaga virilis* (8 hosts) and *Compsilura concinnata* (7 hosts); Braconidae *Protapanteles paleacritae* (7 hosts), *Diolcogaster facetosa* (4 hosts), and *Microplitis hypantriae* (4 hosts); Eulophidae *Eulophus anomocerus* (5 hosts), *E. nebulosus* (5 hosts), and *Euplectrus maculiventris* (4 hosts); and Perilampidae *Perilampus canadensis* (4 hosts). No attempt was made to

determine percentage parasitism contributed by each parasitoid species.

DISCUSSION

The likelihood of rearing multiple species of parasitoids from a given lepidopterous larval species increases as abundance of the larva increases. The two most abundant species of caterpillars collected at Cooper's Rock State Forest in 1984 and 1985 were the noctuids *Polia latex* (Gn.) and *Morrisonia confusa* (Hbn.) (Butler 1992); 16 and 14 species of parasitoids were reared from these two species, respectively. Other abundant species of caterpillars were the geometrids *Itame pustularia* (Gn.) (9 species of parasitoids), *Lomographa vestaliata* (Gn.) (7), *Campaea perlata* (Gn.) (7); the arctiids *Hyphantria cunea* (Drury) (7), and *Halysidota tessellaris* (J. E. Smith) (6); and *Acronica fragilis* (Gn.) (5) and *A. ovata* (Grt.) (5). The geometrids *Melanolophia canadaria* (Gn.) and *Probole amicarica* (H.-S.) were among the most numerous caterpillars, but only four and three species of parasitoids were reared from them, respectively.

During a study of parasitoids associated with an outbreak of a looper complex (Geometridae) in eastern West Virginia in 1983 and 1984, Butler (1990) reared 21 species of parasitoids from *Phigalia titea* (Cram.), the most abundant of the looper species.

Twenty-one of the 136 primary parasitoid/host relationships listed in Table 1, reconfirm records published by Arnaud (1978) for Tachinidae and by Krombein et al. (1979) for hymenopterous taxa. These previous records are, as expected, for lepidopterous hosts which are generally more abundant and/or considered more economically important: *Alsophila pomataria*, *Erannis tiliaria*, *Hydria prunivorata*, *Malacosoma americanum*, *Dryocampa rubicunda*, *Anisota virginiana*, *Nadata gibbosa*, *Hyphantria cunea*, and *Orygia leucostigma*. New primary parasitoid/host records presented here total 115 or 85% of those generated during this study.

Table 1. Parasitoid taxa reared from Macrolepidoptera larvae collected at Cooper's Rock State Forest 1984 and 1985. Secondary parasitoids indicated by "2°."

Macrolepidoptera Species N ¹	Parasitoids	
	Family ²	Genus/Species ³
Drepanidae		
<i>Drepana arcuata</i> Wlk. 2	Brac.	<i>Rogas</i> sp. R
Geometridae		
<i>Alsophila pometaria</i> (Harr.) 9	Brac.	<i>Protapanteles paleacritae</i> (Riley) R
<i>Itame pustularia</i> (Gn.) 234	Tach.	<i>Hyphantrophaga virilis</i> (Aldrich & Webber) U <i>Xanthophyto</i> sp. U <i>Blepharomyia</i> sp. U
	Ichn.	<i>Drusonia deceptor</i> (Walley) U <i>Casinaria forcipata</i> Walley U
	Brac.	<i>Meteorus</i> sp. U <i>Rogas</i> sp. U
	Chal.	<i>Brachymeria aeca</i> Burks (2°)
	Eulo.	<i>Euplectrus maculiventris</i> Westwood U
<i>Glena cribrataria</i> (Gn.) 5	Brac.	<i>Rogas</i> sp. U
<i>Epimecis hortaria</i> (F.) 2	Brac.	<i>Cotesia</i> sp. U
<i>Melanolophia canadaria</i> (Gn.) 228	Tach.	<i>Xanthophyto</i> sp. U
	Ichn.	<i>Aphanistes</i> sp. U
	Brac.	<i>Protapanteles paleacritae</i> (Riley) U <i>Cotesia</i> sp. U
<i>Hypagyrtis unipunctata</i> (Haw.) 11	Pter.	<i>Hypopteromalus tabacum</i> (Fitch) U
<i>Erannis tiliaria</i> (Harr.) 40	Ichn.	<i>Hyposoter fuscitarsis</i> (Vier.) R <i>Phobocampe geometrae</i> (Ash.) R
<i>Lomographa vestaliata</i> (Gn.) 123	Tach.	<i>Compsilura concinnata</i> (Meigen) U <i>Hyphantrophaga virilis</i> (Aldrich & Webber) U
	Ichn.	<i>Aphanistes heinrichi</i> Hopper U <i>Mesochorus pictilis</i> Holmgren (2°)
	Brac.	<i>Protapanteles paleacritae</i> (Riley) U <i>Meteorus</i> sp. U <i>Rogas</i> sp. U
<i>Lomographa glomeraria</i> (Grt.) 183	Tach.	<i>Compsilura concinnata</i> (Meigen) U <i>Hyphantrophaga virilis</i> (Aldrich & Webber) U
	Brac.	<i>Diolcogaster facetosa</i> (Weed) U <i>Meteorus</i> sp. U
<i>Campaea perlata</i> (Gn.) 59	Brac.	<i>Protapanteles paleacritae</i> (Riley) U <i>Diolcogaster facetosa</i> (Weed) U <i>Microgaster</i> sp. U
	Eulo.	<i>Euplectrus maculiventris</i> Westwood U <i>Cirrospilus cinctithorax</i> (Girault) U C. sp. 2 U
	Peri.	<i>Perilampus canadensis</i> Crawford (2°)
<i>Probole amicarua</i> (H.-S.) 237	Ichn.	<i>Mesochorus discitergus</i> (Say) U
	Brac.	<i>Protapanteles paleacritae</i> (Riley) U <i>Cotesia</i> sp. U
<i>Plagodis serinaria</i> (H.-S.) 10	Tach.	<i>Hyphantrophaga virilis</i> (Aldrich & Webber) U <i>Xanthophyto</i> sp. U
	Ichn.	<i>Platylabus hyperetis</i> Heinr. U
<i>Besma endropiaria</i> (G. & R.) 18	Tach.	<i>Hyphantrophaga virilis</i> Aldrich & Webber) U
<i>Nemoria mimosaria</i> (Gn.) 4	Brac.	<i>Protapanteles paleacritae</i> (Riley) U
<i>Hydria prunivorata</i> (Fgn.) 139	Ichn.	<i>Sinophorus hydriae</i> Sanborne U
	Brac.	<i>Cotesia acauda</i> (Prov.) R

Table I. Continued.

Macrolepidoptera Species N ¹	Parasitoids	
	Family ²	Genus/Species ¹
Lasiocampidae		
<i>Malacosoma americanum</i> (F.) 6	Tach.	<i>Compsilura concinnata</i> (Meigen) R
		<i>Lespesia</i> sp. R
	Ichn.	<i>Bathythrix triangularis</i> (Cresson) (2°)
Saturniidae		
<i>Dryocampa rubicunda</i> (F.) 42	Tach.	<i>Eumasicerca sternalis</i> (Coquillett) R
		<i>Lespesia anisotae</i> (Webber) U
<i>Anisota virginiensis</i> (Drury) 71	Ichn.	<i>Hyposoter fugitivus</i> (Say) R
	Ichn.	<i>Hyposoter fugitivus</i> (Say) R
	Trig.	<i>Poecilognalos costalis</i> (Cresson) U
Notodontidae		
<i>Nadata gibbosa</i> (J.E. Smith) 37	Tach.	<i>Compsilura concinnata</i> (Meigen) R
	Ichn.	<i>Ophion flavidus</i> Brullé U
<i>Symmerista leucitys</i> Franc. 46	Tach.	<i>Lespesia stonei</i> Sabrosky U
	Trig.	<i>Poecilognalos costalis</i> (Cresson) U
<i>Macrurucampa marthesia</i> (Cram.) 46	Trig.	<i>Poecilognalos costalis</i> (Cresson) U
<i>Heterocampa guttivitta</i> (Wlk.) 23	Eulo.	<i>Eulophus</i> sp. near <i>koebeleri</i> (Crawford) U
<i>Lochmaeus manteo</i> Doubleday 21	Tach.	<i>Lespesia stonei</i> Sabrosky U
	Brac.	<i>Diolcogaster schizurae</i> (Mues.) U
<i>Schizura unicornis</i> (J.E. Smith) 4	Brac.	<i>Diolcogaster schizurae</i> (Mues.) U
Arctiidae		
<i>Hyphantria cunea</i> (Drury) 73	Tach.	<i>Compsilura concinnata</i> (Meigen) R
		<i>Hyphantrophaga blanda</i> (Osten Sacken) U
		<i>Blondelia hyphantriae</i> (Tothill) R
	Ichn.	<i>Therion sassacus</i> (Vier.) R
	Brac.	<i>Cotesia hyphantriae</i> (Riley) R
		<i>Meteorus hyphantriae</i> (Riley) R
<i>Halysidota tessellaris</i> (J.E. Smith) 63	Eulo.	<i>Elachertus cidariae</i> (Ash.) U
	Tach.	<i>Blondelia hyphantriae</i> (Tothill) R
	Ichn.	<i>Therion morio</i> (F.) U
		<i>Bathythrix triangularis</i> (Cresson) (2°)
		<i>Mesochorus discitergus</i> (Say) (2°)
	Brac.	<i>Cotesia phoberti</i> (Rohwer) R
	Peri.	<i>Perilampus canadensis</i> (Crawford) U
Lymantriidae		
<i>Orygia leucostigma</i> (J.E. Smith) 38	Ichn.	<i>Phobocampe pallipes</i> (Prov.) R
		<i>Iseropus stercorator</i> <i>orygiae</i> (Ash.) R
	Brac.	<i>Meteorus tersus</i> (Mues.) U
	Peri.	<i>Perilampus hyalinus</i> (Say) (2°)
Noctuidae		
<i>Bomolocha baltimoralis</i> (Gn.) 35	Tach.	<i>Oswaldia assimilis</i> (Townsend) U
	Ichn.	<i>Hyposoter annulipes</i> (Cresson) U
	Brac.	<i>Protapanteles paleacritae</i> (Riley) U
<i>Zale minerea</i> (Gn.) 19	Tach.	<i>Xanthophyto</i> sp. Townsend U
	Brac.	<i>Distatrix</i> sp. U
<i>Parallelia bistriaris</i> Hbn. 46	Ichn.	<i>Netelia palpalis</i> (Cush.) U
		<i>Netelia</i> sp. U
	Brac.	<i>Rogas</i> sp. U
<i>Cerma cerintha</i> (Tr.) 7	Tach.	<i>Compsilura concinnata</i> (Meigen) U
<i>Colocasia propinquinelinea</i> (Grt.) 13	Brac.	<i>Microplitis</i> sp. U

Table 1. Continued.

Macrolepidoptera Species N ¹	Parasitoids	
	Family ²	Genus/Species ³
<i>Acronicta americana</i> (Harr.) 19	Brac.	<i>Diolcogaster</i> (poss. <i>facetosa</i>) (Weed) U
<i>Acronicta hasta</i> (Gn.) 14	Tach.	<i>Hyphantrophaga virilis</i> (Aldrich & Webber) U
	Brac.	<i>Rogas</i> sp. U
<i>Acronicta fragilis</i> (Gn.) 124	Tach.	<i>Compsilura concinnata</i> (Meigen) R
	Brac.	<i>Meteorus hyphantriae</i> (Riley) U
		<i>Meteorus communis</i> (Cresson) U
		<i>Meteorus</i> sp. U
		<i>Rogas</i> sp. U
<i>Acronicta ovata</i> (Grt.) 106	Ichn.	<i>Phobocampe</i> n. sp. U
	Brac.	<i>Meteorus hyphantriae</i> (Riley) U
		<i>Meteorus</i> sp. U
	Eulo.	<i>Eulophus anomocerus</i> (Crawford) U
		<i>Eulophus</i> sp. U
<i>Amphipyra pyramidoides</i> (Gn.) 6	Brac.	<i>Microplitis hyphantriae</i> Ash. U
<i>Lithophane hemina</i> (Grt.) 23	Ichn.	<i>Diphyus comes</i> (Cresson) U
	Eulo.	<i>Eulophus anomocerus</i> (Crawford) U
		<i>Eulophus nebulosus</i> (Prov.) U
<i>Eupsila morrisoni</i> (Grt.) 9	Brac.	<i>Microplitis hyphantriae</i> Ash. U
<i>Polia latex</i> (Gn.) 276	Tach.	<i>Hyphantrophaga virilis</i> (Aldrich & Webber) U
		<i>Blondelia hyphantriae</i> (Tothill) U
		<i>Lespesia stonei</i> Sabrosky U
	Ichn.	<i>Hyposoter fugitivus</i> (Say) U
		<i>Alloplasta superba</i> (Prov.) U
		<i>Eutanyacra improvisa</i> (Cresson)
		<i>Mesochorus pictilis</i> Holmgren (2°)
	Brac.	<i>Diolcogaster facetosa</i> (Weed) U
		<i>Microplitis hyphantriae</i> Ash.
		<i>Microplitis</i> sp. U
		<i>Meteorus bakeri</i> C. & D. U
	Eulo.	<i>Euplectrus maculiventris</i> Westwood U
		<i>Euplectrus</i> sp. U
		<i>Eulophus nebulosus</i> (Prov.) U
		<i>Eulophus</i> sp. U
<i>Orthosia rubescens</i> (Wlk.) 6	Peri.	<i>Perilampus canadensis</i> Crawford (2°)
	Eulo.	<i>Eulophus nebulosus</i> (Prov.) U
		<i>Eulophus smerintheta</i> (Ash.) U
		<i>Eulophus anomocerus</i> (Crawford) U
<i>Orthosia hibisci</i> (Gn.) 14	Eulo.	<i>Eulophus nebulosus</i> (Prov.) U
		<i>Eulophus anomocerus</i> (Crawford) U
<i>Crocigrapha normani</i> (Grt.) 6	Eulo.	<i>Eulophus nebulosus</i> (Prov.) U
		<i>Eulophus anomocerus</i> (Crawford) U
<i>Morrisonia confusa</i> (Hbn.) 263	Tach.	<i>Hyphantrophaga virilis</i> (Aldrich & Webber) U
	Ichn.	<i>Hyposoter annulipes</i> (Cresson) U
		<i>Drusonia wyomingensis</i> (Vier.) U
		<i>Enicospilus merdarius</i> (Grav.) U
		<i>Isodromas lycaenae</i> How. U
		<i>Mesochorus vittator</i> (Zett.) U
		<i>Itopectis conquisitor</i> (Say) (2°)
	Brac.	<i>Microplitis hyphantriae</i> Ash. U
		<i>Microplitis</i> sp. U
		<i>Cotesia</i> sp. U

Table 1. Continued.

Macrolepidoptera Species N ¹	Parasitoids	
	Family ²	Genus Species ³
<i>Abagrotis alternata</i> (Grt.) 15	Eulo.	<i>Euplectrus maculiventris</i> Westwood U
		<i>Euplectrus bicolor</i> (Swederus) U
		<i>Pediobius crassicornis</i> (Thomson) U
	Peri.	<i>Perilampus canadensis</i> Crawford (2 ^o)
	Eulo.	<i>Euplectrus bicolor</i> (Swederus) U

¹ Number of specimens reared of each macrolepidoptera species.

² Tach. = Tachinidae; Ichn. = Ichneumonidae; Bracon. = Braconidae; Trig. = Trigonidae; Chal. = Chalcididae; Eulo. = Eulophidae; Pter. = Pteromalidae; Peri. = Perilampidae.

³ R = parasitoid/host relationships recorded by Arnaud (1978) for Tachinidae (Diptera) and Krombein et al. (1979) for hymenopterous species. U = those parasitoid host relationships not recorded in above publications. Secondary parasitoids not considered.

At Cooper's Rock in 1984 and 1985 little defoliation was evident from the 100 species of macrolepidopterous larvae collected on black cherry, black birch, red maple, and mixed oaks. Caterpillar populations were generally low. Among the factors responsible for the low populations were the 74 species of parasitoids obtained in this study. Our knowledge of parasitoid/host relationships for most of our native forest defoliating macrolepidoptera is apparently very limited as evidenced by the large number of new records in this study.

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LITERATURE CITED

- Arnaud, P. H., Jr. 1978. A host-parasite catalog of North American Tachinidae (Diptera). United States Department of Agriculture Miscellaneous Publication No. 1319. 860 pp.

- Butler, L. 1990. Parasitoids of a looper complex (Lepidoptera: Geometridae) in West Virginia. Canadian Entomologist 122: 1041-1043.
- . 1992. The community of macrolepidopterous larvae at Cooper's Rock State Forest, West Virginia: A baseline study. Canadian Entomologist. (In press.)
- Butler, L. and V. Kondo. 1991. Macrolepidoptera moths collected by blacklight trap at Cooper's Rock State Forest, West Virginia: A baseline study. West Virginia Agriculture and Forestry Experiment Station Bulletin 705. 25 pp.
- Carvell, K. L. 1983. A summary of 1973-1982 weather data from the West Virginia University Forest. West Virginia University Forestry Notes 10: 13-16.
- Krombein, K. V. 1958. Hymenoptera of America North of Mexico, Synoptic Catalog, First Supplement. United States Department of Agriculture Monograph 2. 305 pp.
- Krombein, K. V. and B. D. Burks. 1967. Hymenoptera of America North of Mexico, Synoptic Catalog, Second Supplement. United States Department of Agriculture Monograph 2. 584 pp.
- Krombein, K. V., P. D. Hurd, Jr., D. R. Smith, and B. D. Burks. 1979. Catalog of Hymenoptera in America North of Mexico. Vol. 1, Symphyta and Apocrita (Parasitica). Smithsonian Institution Press, Washington, D.C. 1198 pp.
- Muesebeck, C. F. W., K. V. Krombein, and H. K. Townes. 1951. Hymenoptera of America North of Mexico, Synoptic Catalog, United States Department of Agriculture Monograph 2. 1420 pp.
- Raizenne, H. 1952. Forest Lepidoptera of Southern Ontario and Their Parasites Received and Reared at the Ottawa Forest Insect Survey Laboratory from 1937 to 1948. Canada Department of Agricultural

- Science Service, Division of Forest Biology. 277 pp.
- Schaffner, J. V., Jr. and C. L. Griswold. 1934. Macrolepidoptera and their parasites reared from field collections in the northeastern part of the United States. United States Department of Agriculture Miscellaneous Publication 188: 1-160.
- Townes, H. and M. Townes. 1959. Ichneumon-Flies of America North of Mexico: 1. Subfamily Metopiinae. United States Natural History Museum Bulletin 216. 318 pp.
- . 1960. Ichneumon-Flies of America North of Mexico: 2. Subfamilies Ephialtinae, Xoridinae, Acaenitinae. United States Natural History Museum Bulletin 216, Part 2. 676 pp.
- . 1962. Ichneumon-Flies of America North of Mexico: Subfamily Gelinae, Tribe Mesostenini. United States Natural History Museum Bulletin 216, Part 3. 601 pp.
- Viereck, H. L. 1916. Guide to the Insects of Connecticut, Part III, The Hymenoptera, or Wasp-like Insects, of Connecticut. Bulletin No. 22, State Geology and Natural History Survey. 824 pp.